PII: S0957-4166(96)00180-2

Stereoselective Dioxygenase-Catalysed Benzylic Hydroxylation at Prochiral Methylene Groups in the Chemoenzymatic Synthesis of Enantiopure Vicinal Aminoindanols.

Derek R. Boyd,* Narain D. Sharma, Nigel L. Bowers, Peter A. Goodrich, Melanie R. Groocock,

A. John Blacker, David A. Clarke, Tina Howard, and Howard Dalton *

School of Chemistry, The Queen's University of Belfast, Belfast BT9 5AG, UK
 Zeneca PTD, PO Box A38, Leeds Rd, Huddersfield, Yorkshire HD2 1FF, UK
 Department of Biological Sciences, University of Warwick, Coventry CV4 7AL, UK

Abstract: Enantiopure benzylic alcohols containing two stereogenic centres in a cis-relationship result from stereoselective monohydroxylation of achiral 2-substituted indans in cultures of Pseudomonas putida UV4 and are used in the chemoenzymatic synthesis of both cis- and transaminoindanol enantiomers. Copyright © 1996 Elsevier Science Ltd

Previous studies have shown that dioxygenase enzymes present in the soil bacterium *Pseudomonas* putida can catalyse benzylic monohydroxylation¹⁻⁶ involving stereoselective replacement of one prochiral hydrogen atom and creation of a new stereogenic centre in bioproducts of variable enantiopurity. The results now presented differ from the earlier reports¹⁻⁶ since they demonstrate an exclusive stereopreference for one prochiral hydrogen atom and one prochiral methylene group during dioxygenase-catalysed benzylic monohydroxylation with the concomitant creation of two new chiral centres. This phenomenon has been observed during oxidation of the series of achiral 2-substituted indan substrates 1A-1D to yield enantiopure cis-2-methyl-1-indanol 2A, cis-1,2-dihydroxyindan 2B, cis-2-bromo-1-indanol 2C, and cis-2-azido-1-indanol 2D respectively.

Addition of 2-methyindan 1A as substrate to growing shake flask cultures of *P.putida* UV4, using standard conditions previously reported, ^{2.6} gave [1R,2R]-2-methyl-1-indanol (2A, 21% isolated yield, α]_D²⁵ -38, CHCl₃, >98% e.e. by MTPA ester formation; lit. ⁷ α]_D²⁵ +30, CHCl₃). Addition of 2-indanol 1B, 2-bromoindan 1C, and 2-azidoindan 1D as substrates under standard biotransformation conditions in each case yielded the corresponding enantiopure 1-indanols *i.e.* [1S,2R]-2B (50% isolated yield, $[\alpha]_D^{25}$ -48, CHCl₃,

>98% e.e. by MTPA ester formation; lit.⁸ $[\alpha]_D^{25}$ -51, CHCl₃), [1S,2R]-2C (35% isolated yield, $[\alpha]_D^{25}$ -61, CHCl₃, >98% e.e. by MTPA ester formation), [1S,2R]-2D (70% isolated yield, $[\alpha]_D^{25}$ -111, CHCl₃, >98% e.e.; lit.⁹ $[\alpha]_D^{25}$ -111, CHCl₃). The absolute configurations of enantiopure samples of compounds 2A, 2B, and 2D have been reported.⁷⁻⁹ The [1S,2R] absolute configuration assigned to metabolite (-)-2C was based on the stereochemical correlation sequence shown in Scheme 1. The substituted 1-indanol metabolites 2A-2D had the same absolute configuration at C-1 (shown in structure 2) as that found in the parent 1-indanol derived by toluene dioxygenase (TDO)-catalysed hydroxylation of indan ²

Scheme 1

$$NH_2$$
 V_1
 V_2
 V_3
 V_4
 V_4

Reagents (yield): i CH₃CN / H₂SO₄ (70-85%); ii KOH (90%); iii MeSO₂Cl; iv KOH (80%); v NaN₃(70%); vi LiAlH₄ (60%); vii H₂/Pd/C (80%);

Evidence that the monohydroxylation reactions were catalysed by a TDO enzyme was obtained with a recombinant E.coli strain expressing the toluene dioxygenase gene from P.putida NCIMB 11767 on plasmid pKST11. The monol 2A, enantiopurity ca. 91% e.e., of identical absolute configuration and comparable isolated yield (40%) to that obtained using P.putida UV4, was again obtained from biotransformation of substrate 1A using the E.coli pKST11 clone. When the parent E.coli strain was used as a control, the bioproduct 2A was not detected. The mechanism of the TDO-catalysed oxidation is presently unknown, however, the structures of the metabolites obtained when 2-iodoindan 1E was used as substrate with P.putida UV4, were consistent with involvement of an initially formed benzylic radical. Thus, compound 1E was assumed to have been biotransformed, via indene as intermediate, to yield 1R-indenol (8% isolated yield, $[\alpha]_D^{25}$ -249, CHCl₃, >98% e.e.) and cis-1S,2R- dihydroxyindan (2B, $[\alpha]_D^{25}$ -11, CHCl₃, 20% isolated yield, ca.20% e.e.) of identical configuration and enantiopurity to those found as metabolites of indene using P.putida UV4. Since the C-I bond in 1E is weaker than any of the C-R bonds in substrates 1A-1D,

homolytic cleavage of the C-I bond β to a radical centre will be extremely rapid. ¹² It is probable that in compound 1E this homolysis occurs preferentially to yield indene prior to hydroxylation occurring (only traces of the hydroxylation product 2E were found). A similar mechanism involving a carbon centred benzylic radical has recently been postulated for the benzylic hydroxylation of 1,2- and 1,4-dihydronaphthalenes. ^{5,6} Preliminary studies from these laboratories, using specifically labelled precursors, have also shown that the TDO-catalysed benzylic oxidation process occurs with total retention of configuration. ¹³

This enzyme-catalysed synthetic approach to *cis*-diol **2B** *via* benzylic monohydroxylation of 2-indanol **1B**, resulted in high enantiopurity (>98% e.e.) and improved yield (50%) compared with the TDO-catalysed *cis*-dihydroxylation of indene in *P.putida*. ^{1,2,14} Samples of *cis*-diol **2B** of high enantiopurity (\geq 90% e.e.) have been obtained *via* a naphthalene dioxygenase-catalysed (NDO) asymmetric *cis*-dihydroxylation of indene or NDO-catalysed kinetic resolution of racemic *cis*-1,2-dihydroxyindan using *P.Putida* NCIMB 8859. ¹⁴ The ready availability of single enantiomers of the novel bioproducts *cis*-bromohydrin **2C**, and *cis*-hydroxylation process prompted studies of their application in asymmetric synthesis.

Considerable interest has recently been shown in the synthesis ¹⁵⁻¹⁷ and use of enantiopure *cis*-1-amino-2-hydroxyindan 3 and isomers as chiral catalysts in reductions, ^{18,19} in alkyl zinc addition reactions, ²⁰ as chiral ligands in Diels-Alder cycloadditions, ²⁰ and as chiral intermediates in the synthesis of HIV inhibitors. ²¹ The application of enantiopure 1-indanol metabolites 2C and 2D to the synthesis of *cis*-[1S,2R]- and *cis*-[1R,2S]-1-amino-2-hydroxyindan 3, *trans*-[1S,2S]-1-amino-2-hydroxyindan 4, *trans*-[1S,2S]-2-amino-1-hydroxyindan 5 and *cis*-[1S,2R]-2-amino-1-hydroxyindan 6 is shown in Scheme 1.

Using Ritter reaction conditions similar to those reported, 15 for the conversion of epoxide 7 to the cisaminoindanol 3, the enantiopure sample of cis-bromohydrin 2C was converted to cis-[1S,2R]-1-amino-2hydroxyindan 3 ($[\alpha]_{\rm p}^{25}$ -65, CHCl₃; lit. $^{22}[\alpha]_{\rm p}^{25}$ -65, CHCl₃). Similarly the derived [1R,2S]-epoxide 7 ($[\alpha]_{\rm p}^{25}$ -55, CHCl₃; lit. 8 [α] 25 -55, CHCl₃), available via synthesis and inversion of configuration at C-1 of the cisbromomesylate intermediate, was converted into enantiomerically pure cis-[1R,2S]-1-amino-2-hydroxyindan 3 ($[\alpha]_D^{25}$ +65, CHCl₃). Nucleophilic attack of azide at the benzylic C-1 position of the [1R,2S]-epoxide 7 yielded the trans-1-azido-2-hydroxyindan 8 ($[\alpha]_D^{25}$ +75, CHCl₃). The enantiopurity (>98%e.e.) and absolute configuration [1S,2S] of the latter compound was assumed to follow from the precursor [1R,2S]-epoxide 7 and was sterechemically correlated with the derived trans-[1S,2S]-1-amino-2-hydroxyindan 4 ($[\alpha]_n^{25}$ +22, CHCl₃) following LiAlH₄ reduction. Nucleophilic substitution of the bromine atom in the [1S,2R]-cisbromohydrin 2C using sodium azide in DMF gave [18,28]-trans-1-hydroxy-2-azidoindan 9 ($[\alpha]_D^{22}$ +32, CHCl₃; lit. 9 [α] ${}^{25}_{10}$ +32, CHCl₃). Catalytic hydrogenolysis of the hydroxyazides 9 and 2D yielded trans-[18,28]-2-amino-1-hydroxyindan 5 ($[\alpha]_D^{25}$ +16, CHCl₃; lit. $[\alpha]_D^{25}$ +13, CHCl₃) and cis-[18,2R]-2-amino-1hydroxyindan 6 ($[\alpha]_D^{25}$ -63, CHCl₃; lit. $[\alpha]_D^{25}$ -61, CHCl₃) respectively. The aminoindanol enantiomers (+)-3 and (-)-3, (+)-5 and (-)-6 derived from enantiopure metabolites were found to have similar $[\alpha]_D^{25}$ values to those previously synthesised by alternative methods, and are assumed to have the reported 9,22 absolute configurations. In conclusion, stereoselective TDO-catalysed benzylic hydroxylation of 2-substituted indans allied to chemical synthesis has provided a new chemoenzymatic route to the corresponding cis-11562 D. R. BOYD et al.

hydroxyindan enantiomers which have been utilized in the synthesis of a range of enantiopure vicinal aminoalcohols.

Acknowlegements: We thank Professor T.P.Begley (Cornell University) for helpful discussion and the following bodies for financial support: BBSRC (to NDS, DAC, and TH), IRTU (to MRG), DENI for a Distinction Award (to NIB) and a CAST Award with Zeneca PTD (to PAG).

References

- 1. Wackett, L.P.; Kwart L.D.; Gibson, D.T. Biochemistry 1988, 27, 1360-1367.
- Boyd, D.R.; Sharma, N.D.; Stevenson, P.J.; Chima, J.; Dalton, H. Tetrahedron Lett. 1991, 32, 3887-3890.
- 3. Resnick, S.M.; Torok D.S.; Lee, K.; Brand, J.M.; Gibson, D.T. Appl. Environ. Microbiol. 1994, 60, 3323-3328.
- 4. Gibson, D.T.; Resnick, S.M.; Lee, J.M.; Brand, J.M.; Torok, D.S. J.Bacteriol. 1995, 177, 2615-2621.
- 5. Torok, D.S.; Resnick, SM.; Brand, J.M.; Cruden, D.L.; Gibson, D.T. J. Bacteriol. 1995, 177,5799-5805.
- 6. Boyd, D.R.; Sharma, N.D.; Dalton, H.; Kerley, N.A.; McMordie, R.A.S.Sheldrake, G.N.; Williams, P.; J. Chem. Soc. Perkin Trans 1. 1996, 64-74.
- 7. Jaouen, G; Meyer, A. J.Am. Chem. Soc. 1975, 97, 4667-4672.
- 8. Boyd, D.R.; Sharma, N.D.; Smith, A.E. J. Chem. Soc. Perkin Trans. 1. 1982, 2767-2770.
- 9. Mitrochkine, A.; Gil, G., Reglier; M. Tetrahedron: Asymmetry 1995, 6, 1535-1538.
- Allen, C.C.R.; Boyd, D.R.; Dalton, H.; Sharma, N.D.; Haughey, S.A.; McMordie, R.A.S.; McMurray, B.T.; Sheldrake, G.N.; Sproule K. J. Chem. Soc. Chem. Commun. 1995, 119-120
- 11. Boyd, D.R.; McMordie, R.A.S.; Sharma, N.D.; Dalton, H.; Williams, P.; Jenkins, R.O. J. Chem. Soc. Chem. Commun. 1989, 339-340.
- 12. Burdi, D.; Begley, T.P. J.Am. Chem. Soc. 1991, 113, 7768-7770.
- 13. Boyd, D.R.; Sharma, N.D.; Dalton, H.; in 'Organic Reactivity: Physical and Biological Aspects', Eds. Golding, B.T.; Griffin, R.J.; Maskill, H.; RSC, Cambridge, 1995, 130-139.
- Allen, C.C.R.; Boyd, D.R.; Sharma, N.D.; Dalton, H.; Brannigan, I.; Kerley, N.A.; Sheldrake, G.N.;
 Taylor, S.C. J. Chem. Soc. Chem. Commun. 1995, 117-119.
- Senanayake, C.H.; Roberts, F.E.; DiMichele, L.M.; Ryan, K.M.; Liu, J.; Fredenburgh, L.E.Foster, B.S.;
 Douglas, A.W.; Larsen, R.D.; Verhoeven, T.R.; Reider, P.J. Tetrahedron Lett. 1995, 36, 3993-3996.
- Senanayake, C.H.; DiMichele, L.M.; Liu, J.; Fredenburgh, L.E.; Ryan, K.M.; Roberts, F.E.; Larsen R.D.; Verhoeven, T.R.; Reider, P.J. Tetrahedron Lett. 1995, 36, 7615-7618.
- 17. Mitrochkine, A.; Eydoux, F.; Martres, M.; Gil, G., Heumann, A.; Reglier, M. Tetrahedron: Asymmetry 1995, 6, 59-62.
- 18. DiSimone, B.; Savoia, D.; Tagliavini, E.; Umani-Ronchi, A. Tetrahedron Asymmetry 1995, 6,301-306.
- 19. Ghosh, A.K.; Chen, Y.; Tetrahedron Lett. 1995, 36, 6811-6814.
- 20. Davies, I.W.; Senanayake, C.H.; Castonguay, L.; Larson, R.D.; Verhoeven, T.R.; Reider, P.J.; Tetrahedron Lett. 1995, 36, 7619-7622.
- Lyle, T.A.; Wishcount, C.M.; Grace, J.P.; Thompson, W.J.; Anderson P.S.; Darke, P.L.; Zugay, J.A.;
 Emini, E.A.; Scheif, W.A.; Quintero, J.C.; Dixon, R.A.; Sigal, I.S.; Huff, J.R. J.Med.Chem. 1991, 34, 1228-1230.
- 22. Didier, E.; Loubinoux, B.; Rihs, G. Tetrahedron 1991, 47, 4941-4958.